

# Connor J. Kenny, PhD

## Scientist, RNA Therapeutics & Computational Genomics

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Scientist combining computational genomics, assay development, and high-throughput functional validation to identify disease mechanisms and therapeutic opportunities. Builds interpretable models from large genomics datasets and translates sequence-level insights into experimentally testable mechanisms, with a publication record linking RNA mechanisms, human genetics, and cancer biology.

## SELECTED IMPACT

- **First-author mechanism paper:** defined LUC7-regulated 5' splice-site classes in *Nature Communications*.
- **Therapeutic relevance:** identified leukemia/MDS splice signatures linked to splicing-modulator sensitivity.
- **Platform scale:** built pipelines spanning 2,500+ RNA-seq datasets and ~14,000 clinical DNA samples.
- **Deep-learning platform development:** experience with building interpretable deep learning models for RNA sequence design.

## TECHNICAL TOOLBOX

**Computational:** Python, R, Bash; Snakemake; ML/Bayesian modeling; SLURM/HPC; genomics database curation

**Experimental:** robotic liquid handling, platform development and optimization, reporter design; cell culture

**Translational:** clinical genomics; patient cohort analysis; mechanism-of-action studies; SOPs

**Disease areas:** RNA biology; neurodevelopmental genetics; myeloid neoplasms/leukemia; cancer genomics

## RESEARCH EXPERIENCE

### University of Chicago | Yang I. Li Lab

Section for Precision Medicine, Chicago, IL

### Postdoctoral Scholar

Sep 2025 - Present

#### Therapeutic targeting of pre-mRNA splicing

- **Therapeutic modeling:** built interpretable convolutional neural networks for *de novo* learning of sequence determinants of splicing-modulating ASOs, small molecules, and cancer-associated non-coding RNAs.
- **Large-scale genomics:** map splicing-factor loss/mutation to stage-specific error signatures across 2,500+ RNA-seq datasets.
- **Statistical modeling:** designed Bayesian models to identify sequence features linked to splicing-modulator activity.

### Massachusetts Institute of Technology | Christopher B. Burge Lab

Department of Biology, Cambridge, MA

### Graduate Student

Sep 2020 - Aug 2025

#### Quantitative modeling of 5' splice site subclass regulation and evolution

- **First-author discovery:** defined LUC7-regulated 5' splice-site classes in *Nature Communications*.
- **Wet/dry validation:** combined RNA-seq, motif modeling, splicing reporter design/cloning, and cross-species validation.
- **Therapeutic relevance:** identified leukemia/MDS cohorts with signatures linked to splicing-modulator sensitivity.

### Boston Children's Hospital / HHMI | Christopher A. Walsh Lab

Division of Genetics and Genomics, Boston, MA

### Research Assistant II

June 2016 - May 2019

#### Rare disease genetics, functional genomics, and neurodevelopmental model systems

- **NGS scale-up:** established robotic liquid-handling protocols for targeted sequencing of ~14,000 human clinical DNA samples.
- **Variant validation:** partnered with clinicians and genetic counselors to validate rare-disease mutations.
- **Functional genomics:** performed transcriptome analysis and devised assays for rare human brain cell types.
- **Team enablement:** created SOPs and accessible analysis tools for somatic and germline variant validation.

## THERAPEUTIC TARGET CONSULTING

**Independent consultant:** assessed therapeutic viability of *SCN1A* poison exon modulation for treating genetic epilepsy.

## TECHNICAL METHODS

**Genomics analysis:** bulk RNA-seq, single-cell transcriptomics, alternative splicing analysis, targeted DNA-seq, eCLIP interpretation, public cohort analysis, and human variant interpretation.

**Modeling and statistics:** machine learning, Bayesian modeling, probabilistic sequence models, statistical analysis, data visualization, and scalable Snakemake/HPC workflows.

**Experimental biology:** cell culture, minigene/splicing reporter design and cloning, RT-PCR/ddPCR, NGS library preparation, FACS/flow sorting, robotic liquid handling, and fluorescence microscopy.

**Research operations:** SOP development, reproducible pipelines, technical documentation, mentorship, manuscript writing, and collaboration across computational and experimental teams.

## EDUCATION

**Massachusetts Institute of Technology** | 2025

PhD, Biology; PhD Presidential Fellow

**Boston University, College of Arts and Sciences** | 2016

BA, Neuroscience with Honors; magna cum laude; GPA 3.83

## SELECTED PUBLICATIONS

**14 peer-reviewed publications; selected examples:**

**Kenny, C.J. et al.** LUC7 proteins define two major classes of 5' splice sites in animals and plants. *Nature Communications*, 2025. Featured in MIT News: MIT biologists discover a new type of control over RNA splicing.

**Shin, T. et al.** Rare variation in non-coding regions with evolutionary signatures contributes to autism risk. *Cell Genomics*, 2024.

**McDonough, G. et al.** Neuropathologically directed profiling of *PRNP* somatic and germline variants in sporadic human prion disease. *Acta Neuropathologica*, 2024.

**Kodani, A., Kenny, C.J. et al.** Posterior neocortex-specific regulation of neuronal migration by *CEP85L* identifies maternal centriole-dependent activation of CDK5. *Neuron*, 2020.

**Smith, R.S., Kenny, C.J. et al.** Sodium Channel *SCN3A* regulation of human cerebral cortical folding and oral motor development. *Neuron*, 2018.

**Ganz, J. et al.** Rates and patterns of clonal oncogenic mutations in normal human brain. *Cancer Discovery*, 2022.

## AWARDS & RECOGNITION

**RNA Canada ARN Conference for RNA Therapeutics:** Top 5% Poster Prize.

**RNA Society:** Selected speaker.

**MIT:** Praecis Presidential Fellowship.

**Boston Children's Hospital:** SPOT Award for outstanding employees.

## LEADERSHIP & COLLABORATION

**Scientific mentorship:** supervised undergraduate, graduate, and visiting PhD researchers on computational biology, machine learning, probabilistic modeling, and evolutionary genomics projects.

**Cross-functional work:** extensive experience collaborating with clinical, computational, and experimental teams.

**Communication:** translated statistical and genomics analyses into assay plans, SOPs, presentations, and manuscript figures.